Organic & Biomolecular **Chemistry**

Cite this: Org. Biomol. Chem., 2012, **10**, 1381

[Dynamic Article Links](http://dx.doi.org/10.1039/c1ob05875d) (

Copper-catalyzed synthesis of substituted indazoles from 2-chloroarenes at low catalyst-loading†

Shinji Tanimori,* Yasuyuki Kobayashi, Yasukazu Iesaki, Yuka Ozaki and Mitsunori Kirihata

Received 2nd June 2011, Accepted 8th November 2011 **DOI: 10.1039/c1ob05875d**

An efficient and convenient access to 1-substituted indazol-3-ones **2** has been achieved throughout the intramolecular C–N bond formations of 2-chloro-benzoic acid-*N*¢-aryl and alkyl-hydrazides employing 0.5 mol% of cuprous (I) iodide and 20 mol% of L-proline as catalyst precursors under mild conditions in moderate to excellent yields.

Introduction

Substituted indazol-3-ones are present in many important pharmaceutically active molecules.**¹** For instance, the indazole compounds and their derivatives have been reported to exhibit analgesic,**²** antitumor,**³** anticancer,**⁴** anti-inflammatory,**⁵** and antifertility activities.**⁶** We previously reported the synthesis of 1 substituted indazol-3-ones from 2-iodo- and 2-bromo-benzoic acid-*N*^{\prime}-aryl and alkylhydrazides, which employed 10 mol% of cuprous(I) iodide and 20 mol% of L-proline as the pre-catalysts at room temperature for 3 h in good to excellent yields (Scheme 1).**7,8** However, decreasing the catalyst-loading under 10 mol% and also the use of chloroarenes as substrates afforded unsatisfactory results.**⁷ Comparison Companies Content Companies (Angers of Companies and Companies Companies and Companies and Companies and Comp**

Scheme 1 Strategy to synthesize indazolones.

The employment of 2-chlorobenzohydrazides as starting materials would represent some benefits. Economically, 2-chlorobenzoic acids and derivatives, precursors of 2-chlorobenzohydrazides, are usually less expensive than the corresponding iodo- and bromoderivatives. Moreover, the rich existence of their analogues with a variety of substituents should also be attractive for diversityoriented synthesis. Ordinarily, iodo- and bromoarenes have been utilized as coupling partner for the copper-catalyzed carboncarbon and carbon-heteroatom bond-forming reactions and choroarenes were not nearly employed due to their significantly low reactivity.**⁹** Moreover, lowering the catalyst loading is also highly desirable from a green chemistry viewpoint. Mostly, 5 to 20 mol% of catalysts have been frequently exploited for the complete consumption of starting materials, in the copper-catalyzed systems.**⁹** After the several examinations based on our previous report,**⁷** it was eventually found that the 2-chlorobenzohydrazides **1** were also suitable substrates to be converted smoothly into 1 substituted indazol-3-ones **2** with only 0.5 mol% of copper catalyst in practical efficiency. Details of the results are reported below.

Results and discussions

Synthesis

Table 1 shows the summary for optimization studies of the reaction conditions to improve our previous system.**⁷** The reaction of 2-chlorobenzoic acid-*N*¢-phenylhydrazide **1a** in the presence of 1 mol% of cuprous (I) iodide with L-proline**9d** (2 mol%)**¹⁰** and potassium carbonate (2 equiv.) in dimethylsulfoxide at 70 *◦*C for 15 h gave a ring-closed product **2a** in 31% yield (entry 1). The reaction did not perform under ligand free conditions (not shown). Gratifyingly, when 5 mol% L-proline was employed with 1 mol% of cuprous (I) iodide, the yield was increased to 67% in the same reaction conditions (entry 2). To our delight, the satisfactory results were observed when excess amount of L-proline (10 and 20 mol% based on substrates) was introduced with 1 mol % CuI to afford desired product in 75 and 79% yields, respectively (entry 3 and 4). Organic base DBU should also be a good base for this transformation, although prolonged reaction time was required to accomplish the complete consumption of the starting material (entry 5). The reaction with cesium carbonate afforded a lower yield (62%, entry 7) and sodium *t*-butoxide provided complex mixture (not shown). A more efficient and economical conversion was realized with the use of 0.5 mol% copper catalyst and large excess L-proline (20 mol%) to produce indazolone **2a** in 80% yield after 24 h at 90 *◦*C (entry 8).**¹¹** To our knowledge, these results would be the first cases for copper-mediated coupling reactions to achieve efficient and practical carbon-heteroatom bond formations utilizing less reactive chloroarene as coupling partner along with a lower catalyst loading $\left($ < 1 mol%) under the mild conditions. The role of excess L-proline based on the loading

Department of Bioscience and Informatics, Graduate School of Life and Environmental Sciences, Osaka Prefecture University, 1-1 Gakuencho, Nakaku, Sakai, Osaka, 599-8531, Japan. E-mail: tanimori@bioinfo.osakafu-u.ac.jp; Fax: +81 72 254 9469; Tel: +81 72 254 9469

[†] Electronic supplementary information (ESI) available: See DOI: 10.1039/c1ob05875d

Table 1 The intramolecular cyclization of **1a** under various reaction conditions*^a*

^a All of the reactions were carried out using benzhydrazide **1a** (1.0 mmol), catalyst (0.5 and 1 mmol), ligand (2–20 mmol), and base (2.0 mmol) in DMSO (10 mL). *^b* Isolated yield.

of copper catalyst for this system could be explained by considering the competitive role of the hydrazide moiety in complexating the copper ion. Such a complexation would inhibit the intramolecular coupling. Therefore a large excess of proline is needed to ensure the complexation of copper. The closely related phenomenon has been argued in our previous report, where DMEDA (20 mol%) and cuprous (I) chloride (1 mol%) were exploited as pre-catalysts for the coupling of 2-haloaniline with α –amino acids to form quinoxaline-2-ones.**¹²** A control experiment without the use of catalyst gave no desired product (not shown).

As the optimized reaction conditions were in hand, we next investigated the cyclization reactions with a series of substituted 2-chlorobenzohydrazides to examine the generality of the conditions (Table 2). Both electron-donating and electron-withdrawing groups on aromatic rings of corresponding benzoic acid and hydrazine moieties are feasible for this cyclization reaction. Alkylsubstituted hydrazides also reacted effectively at the same procedure (entries 7, 8 and 10). Although nitro-substituted hydrazides **1c** and **1d** provided complex mixtures under the optimized conditions, to our delight, the cyclization proceeded smoothly at room temperature to afford indazolone **2c** and **2d** in 93% and 55% yields, respectively (entries 3 and 4). In the latter case, a strong electron-donating methoxy group would destabilize the intermediacy nucleophilic nitrogen anion in the cyclization step. The pyridine containing substrates **1k**, **1l** and **1m** were also accessible for this transformation to provide a novel class of pyrazolo[3,4-b]pyridin-3-ones **2k**, **2l** and **2m** in moderate to good yields (entries 11, 12 and 13). Sulfonamide **1n** also reacted to give multi-substituted indazolone **2n** without the use of protecting group. Moreover, the comparable efficiencies were observed by a comparison of Br/I and Cl as leaving groups as shown in entries 1 and 7. The scalability of this transformation has also been demonstrated with the use of hydrazide **1a** (1.30 g, 5.25 mmol) to provide indazolone **2a** (1.00 g, 90%) under the optimal conditions (see experimental section).

As shown in Scheme 2, indazolone **2o**, **¹³** a key intermediate for the synthesis of (*S*)-5-fluoro-1-(2-fluorophenyl)-3-(piperidin-3-ylmethoxy)-1*H*-indazole **3**, a norepinephrine/serotonin reuptake inhibitor for the treatment of fibromyalgia,**¹⁴** has been

Scheme 2 Simple synthesis of the key intermediates for a pharmaceutical.

synthesized easily from commercial starting materials utilizing our protocol. The copper-mediated cyclization of hydrazides **1o**, derived from 5-fluoro-2-bromo and chlorobenzoic acids with 2 fluorophenylhydrazine, underwent to form desired indazolone **2o** in both 58% yields. The moderate efficiency for the cyclization step would attribute to steric bulk of *ortho*-substitution on hydrazine aromatic ring. This strategy avoids a hazardous reagent and unstable intermediate required for previous route.**¹³**

Pharmacology

In the course of our continuing study directed toward the discovery of anti-cancer drugs,**¹⁵** we tested the cytotoxic activity of all synthesized compounds (hydrazides **1** and indazolones **2**) against HeLaS3 cell line. Most of compounds did not show significant inhibition under 30 μ M (IC₅₀) except for hydrazides **1n** (0.25 μ M) and $10-Br$ (2.9 μ M). Other biological studies are now under investigation.

Conclusions

In conclusion, we have established a general and practical access to substituted indazolones from readily available starting materials. The substrate scope for this synthesis has been expanded to even less reactive chloroarenes in this study. The improved reaction

^a All the reactions were carried out using hydrazide **1** (1.0 mmol), CuI (0.0050 mmol, 0.5 mol%), L-proline (0.020 mmol, 20 mol%), K₂CO₃ (2.0 mmol) in DMSO (10 mL). *^b* Isolated yield. *^c* The reaction was performed at room temperature.

conditions with lower copper catalyst loading (0.5 mol) have been realized by carefully controlled ligand loading (20 mol%). The success of the key low loading of copper-catalyst in this transformation would partially attributed to the use of excess ligand. Bolm suggests that the ligand shifts equilibria away from favorable low-coordinated copper species which otherwise would be deactivated by aggregation.**¹²** Structurally diverse indazolones together with benzhydrazides as the precursors have been synthesized efficiently by this process to prepare two series of small chemical libraries. The simple synthesis of a key intermediate **2o** for a drug **3** was conducted by this protocol in mild conditions. Finally, the novel class of strong cytotoxic compound **1n** has appeared among the libraries.

Experimental

Chemistry

General methods. Unless stated otherwise, reactions were monitored by thin layer chromatography (TLC) on silica gel plates (60 F_{254}), visualizing with ultraviolet light or iodine spray. Column chromatography was performed on silica gel (60–120 mesh) using hexane and ethyl acetate. ¹H and ¹³C NMR spectra were determined in DMSO-d6 solution using 400 and 100 MHz spectrometers, respectively. Proton chemical shifts (δ) are relative to tetramethylsilane (TMS, $\delta = 0.0$) as the internal standard and expressed in parts per million. Spin multiplicities are given as s (singlet), d (doublet), t (triplet), and m (multiplet) as well as b (broad). Coupling constants (*J*) are given in hertz. Infrared spectra were recorded on a FT-IR spectrometer. Melting points were determined by using a Büchi melting point B-540 apparatus and are uncorrected. MS spectra were obtained on a mass spectrometer. HR-MS was determined using JEOL JNM-AX 500 mass spectrometer.

General procedure for the preparation of acid hydrazides 1a–o by the condensation of acid with hydrazine. To a stirred solution of acid (5.0 mmol) in DMF (10 mL) were added EDC·HCl (1.1 g, 5.5 mmol), HOBt·H2O (0.74 g, 5.5 mmol), DMAP (31 mg, 0.25 mmol), and hydrazine (5.0 mmol) at 0 *◦*C. The resulting mixture was allowed to warm to room temperature over 24 h, and it was diluted with EtOAc and poured into saturated NH4Cl solution. Extractive workup with EtOAc and purification by column chromatography afforded acid hydrzide as a white solid.

Spectral data of acid hydrazides (1a–o)

2-Chloro-*N*¢**-phenylbenzohydrazide (1a)⁷ .** Yield 90%; *R*^f 0.53 (hexane : AcOEt = 1 : 1); mp 152.2 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) δ 10.21 (d, 1H, $J = 2.7$ Hz, CONH), 8.03 (d, 1H, *J* = 2.7 Hz, PhNH), 7.38–7.60 (m, 4H, Ar), 7.17 (dd, 2H, *J* = 7.3, 7.6 Hz, Ar), 6.84 (d, 2H, *J* = 7.6 Hz, Ar), 6.72 (t, 1H, *J* = 7.3 Hz, Ar).

2-Chloro-4-methyl-benzoic acid *N*¢**-phenyl-hydrazide (1b).** Yield 68%; mp 178.4 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.14 (s, 1H, CONH), 7.96 (s, 1H), 7.41 (d, 1H, *J* = 8.3 Hz), 7.35 (s, 1H), 7.31 (d, 1H, *J* = 8.0 Hz), 7.17 (t, 2H, *J* = 7.3 Hz), 6.86 (d, 2H, *J* = 7.8 Hz), 6.73 (t, 1H, *J* = 7.3 Hz), 2.35 (s, 3H, Me); ¹³C NMR (136 Hz, DMSO-d6)*d* 166.3, 149.1, 136.8, 135.0, 131.7, 129.5, 129.4, 128.7 (2×C), 127.2, 118.6, 112.3 (2×C), 20.4; HRMS-EI calcd for $C_{14}H_{13}N_2$ OCl: 260.0717, Found: 260.0699.

2-Chloro-4-nitro-benzoic acid *N*¢**-phenyl-hydrazide (1c)¹⁶.** Yield 68%; *R*_f 0.63 (hexane : AcOEt = 1 : 1); mp 169.1 °C; ¹H NMR (400 MHz, DMSO-d6) *d* 10.53 (s, 1H, CONH), 8.46 (d, 1H, *J* = 1.7 Hz, Ar), 8.34 (d, 1H, *J* = 8.5 Hz, Ar), 8.19(s, 1H, PhNH), 7.90 (d, 2H, *J* = 8.5 Hz, Ar), 7.25 (t, 2H, *J* = 7.8 Hz, Ar), 6.92 (d, 2H, *J* = 7.8 Hz, Ar), 6.81 (t, 1H, *J* = 6.8 Hz, Ar); 13C NMR (136 Hz, DMSO-d6) δ 164.9 (C=O), 148.8, 148.6, 141.0, 131.5, 130.4, 128.9 (2¥C, Ar), 124.8, 122.5, 118.9, 112.4 (2¥C, Ar).

2-Chloro-4-nitro-benzoic acid *N*¢**-4-methoxyphenyl-hydrazide (1d).** Yield 59%; R_f 0.54 (hexane : AcOEt = 1 : 1); mp: 183.4 °C; ¹H NMR (400 MHz, DMSO-d6) *δ* 10.42 (s, 1H, CONH), 8.39 (d, 1H, *J* = 2.0 Hz, ArNH), 8.27 (dd, *J* = 6.0, 2.2 Hz, 1H, Ar), 7.78– 7.82 (m, 2H, Ar), 6.78–6.88 (m, 4H, Ar), 3.67 (s, 3H, MeO); 13C NMR (136 Hz, DMSO-d6) δ 164.9 (C=O), 152.9, 148.5, 142.6, 141.1, 131.4, 130.4, 124.7, 122.5, 114.3 (2¥C, Ar), 113.9 (2¥C, Ar), 55.3; IR (neat): 3256 (NH), 2837 (Me), 1660 (C=O), 1518, 1351 (NO₂), 1239, 1037, 832 cm⁻¹; HRMS Calcd for C₁₄H₁₂O₄N₃Cl: 321.0516, Found: 321.0338.

2,4-Dichloro-benzoic acid *N*¢**-phenyl-hydrazide (1e)¹⁷.** Yield 84%; mp 169.1 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.26 (s, 1H, CONH), 8.03 (s, 1H), 7.75 (d, 1H, *J* = 1.7 Hz), 7.52–7.62 (m, 2H), 7.17 (t, 2H, *J* = 7.6 Hz), 6.84 (d, 2H, *J* = 7.6 Hz), 6.73 (t, 1H, *J* = 7.3 Hz); 13C NMR (136 Hz, DMSO-d6) *d* 165.5, 149.0, 135.0, 134.1, 131.5, 130.6, 129.4, 128.8 (2¥C), 127.5, 118.8, 112.3 (2¥C).

2,4-Dichloro-benzoic acid *N*¢**-(4-nitro-phenyl)-hydrazide (1f)¹⁸.** Yield 84%; *R*^f 0.43 (hexane : AcOEt = 1 : 1); mp 247.6 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.63 (s, 1H, CONH), 9.35 (s, 1H, Ar), 8.10 (d, 2H, *J* = 7.8 Hz,Ar), 7.79 (d, 1H, *J* = 1.7 Hz, ArNH), 7.66 (d, 1H, *J* = 8.3 Hz, Ar), 7.57 (dd, 1H, *J* = 1.7, 8.3 Hz, Ar), 6.90 (d, 2H, $J = 7.8$ Hz, Ar); ¹³C NMR (136 Hz, DMSO-d6) δ 165.4 (C=O), 154.5, 138.3 (2×C, Ar), 135.5, 133.4, 131.6, 130.7, 129.5, 127.6, 126.0 (2¥C, Ar), 110.9.

2,4-Dichloro-benzoic acid *N*¢**-***t***-butyl-hydrazide (1g)¹⁹.** Yield 91%; mp 112.9 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 9.80 (br s, 1H), 7.70 (d, 2H, *J* = 2.0 Hz), 7.49 (dd, 1H, *J* = 2.0, 8.3 Hz), 7.44 (d, 1H, $J = 8.3$ Hz), 1.06 (s, 9H); ¹³C NMR (136 Hz, DMSO-d6) δ 164.2, 134.6, 134.5, 131.5, 130.7, 129.1, 127.3, 54.7, 27.3 (3¥C).

2,4-Dichloro-benzoic acid *N*¢**-cyclohexyl-hydrazide (1h).** Yield 60%; mp 164.7 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 9.83 (d, 1H, *J* = 2.7 Hz, CONH), 7.68 (d, 1H, *J* = 1.7 Hz), 7.48 (dd, 1H, *J* = 1.7, 8.3 Hz), 7.43 (1H, d, *J* = 8.3 Hz), 4.94 (br s, 1H), 2.72–2.84 (m, 1H), 1.83 (d, 2H, *J* = 11.5 Hz), 1.63–1.76 (m, 2H), 1.50– 1.59 (d, 2H, *J* = 8.8 Hz), 1.02–1.28 (m, 5H); 13C NMR (136 Hz, DMSO-d6) *d* 164.3, 134.6, 134.5, 131.5, 130.5, 129.1, 127.3, 57.8,

30.9 (2×C), 25.7, 23.9 (2×C); HRMS Calcd for $C_{13}H_{16}N_2OCl_2$: 286.0640, Found: 286.0640.

2-Chloro-5-fluoro-benzoic acid *N*¢**-4-methylphenyl-hydrazide (1i).** Yield 69%; mp 167.0 *◦*C; ¹ H NMR (400 MHz, DMSOd6) *d* 10.25 (d, 1H, *J* = 2.9 Hz), 7.85 (d, 1H, *J* = 2.4 Hz), 7.60 (dd, 1H, *J* = 4.9, 9.0 Hz), 7.44 (dd, 1H, *J* = 2.9, 8.3 Hz), 7.38 (1H, dt, *J* = 2.9, 8.3 Hz), 6.98 (d, 2H, *J* = 8.3 Hz), 6.77 (d, 2H, *J* = 8.3 Hz), 2.19 (s, 3H, Me); 13C NMR (136 Hz, DMSO-d6) *d* 155.8, 137.6, 136.2, 134.4, 129.9 (3C), 120.8 (3¥C), 117.3, 111.9, 104.5, 20.5; HRMS Calcd for C₁₄H₁₂N₂OClF: 278.0622, Found: 278.0605.

2-Chloro-5-fluoro-benzoic acid *N*¢**-***t***-butyl-hydrazide (1j).** Yield 79%; mp 103.3 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 9.91 (br s, 1H), 7.63 (dd, 1H, *J* = 4.9, 8.3 Hz), 7.36–7.46 (m, 2H), 4.93 (br s, 1H), 1.14 (s, 9H); ¹³C NMR (136 Hz, DMSO-d6) δ 163.8 (C=O), 161.6 (159.1), 137.34 (137.26), 131.6 (131.5), 125.72 (125.70), 118.0 (117.8), 116.5 (116.2), 54.7, 27.3 (3¥C); HRMS Calcd for C₁₁H₁₄N₂OClF: 244.0779, Found: 244.0829.

2,6-Dichloro-nicotinic acid *N*¢**-phenyl-hydrazide (1k).** Yield 93%; *R*_f 0.45 (hexane : AcOEt = 1 : 1); mp 147.4. [°]C; ¹H NMR (400 MHz, DMSO-d6) *d* 10.40 (s, 1H, CONH), 8.12 (d, 1H, *J* = 8.0 Hz, Ar), 8.09 (br s, 1H, ArNH), 7.71 (d, 1H, *J* = 8.0 Hz, Ar), 7.17 (t, 2H, *J* = 7.6 Hz, Ar), 6.85 (d, 2H, *J* = 7.8 Hz, Ar), 6.68–6.80 (m, 1H, Ar); 13C NMR (136 Hz, DMSO-d6)*d* 164.2 $(C=0)$, 149.8, 148.7, 146.2, 141.6, 130.7, 128.9 (2×C, Ar), 123.8, 119.0, 112.4 (2¥C, Ar); IR (neat): 3256 (NH), 3055 (Ar), 1653 (C=O), 1492, 1341, 1144, 912, 756, 691 cm⁻¹; HRMS Calcd for $C_{12}H_9N_3OCl_2$: 281.0122, Found: 281.0216. DMSO-49 β 0.021 of 1H, $J = 2.7$ Hz, CONH, 8.03 (d. 1H, 30 (d. 2H, 32 (d

2,6-Dichloro-nicotinic acid *N*¢**-4-chlorophenyl-hydrazide (1l).** Yield 77%; mp 180.3 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.45 (s, 1H, CONH), 8.13 (dd, 1H, *J* = 1.5, 8.0 Hz), 7.71 (dd, 1H, *J* = 1.5, 8.0 Hz), 7.21 (d, 2H, *J* = 7.6 Hz), 6.85 (d, 2H, *J* = 7.6 Hz); ¹³C NMR (136 Hz, DMSO-d6) δ 164.2, 149.9, 147.7, 146.2, 141.6, 130.5, 128.7 (2¥C), 123.8, 122.3, 113.9 (2¥C); HRMS Calcd for C12H8N3OCl3: 314.9733, Found: 314.9837.

2,6-Dichloro-nicotinic acid *N*¢**-4-methylphenyl-hydrazide (1m).** Yield 78%; mp 145.5 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.34 (d, 1H, *J* = 2.4 Hz, CONH), 8.09 (d, 1H, *J* = 8.0 Hz), 7.91 (br s, 1H), 7.70 (d, 1H, *J* = 8.0 Hz), 6.99 (d, 2H, *J* = 8.5 Hz), 6.76 (d, 2H, *J* = 8.5 Hz), 2.18 (s, 3H, Me); 13C NMR (136 Hz, DMSO-d6) *d* 164.1, 149.7, 146.4, 146.2, 141.6, 130.7, 129.2 (2¥C), 127.6, 123.7, 112.6 (2×C), 20.2; HRMS Calcd for $C_{13}H_{11}N_3OCl_2$: 295.0279, Found: 295.0259.

2,4-Dichloro-5-sulfamoyl-benzoic acid *N*¢**-(1-methyl-phenyl) hydrazide (1n).** Yield 71%; R_f 0.40 (hexane : AcOEt = 1 : 1); mp 208.3 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d*: 10.39 (d, 1H, *J* = 2.2 Hz, CONH), 8.00 (s, 1H, Ar), 7.99 (s, 1H, Ar), 7.92 (br s, 1H, ArNH), 7.89 (br s, 2H, SO_2NH_{2} , 6.99 (d, 2H, $J = 8.0$ Hz, Ar), 6.73 (d, 2H, *J* = 8.0 Hz, Ar), 2.18 (s, 3H, Me); 13C NMR (136 Hz, DMSO-d6) δ 164.5 (C=O), 146.5, 140.1, 134.4, 134.2, 132.4, 132.3, 129.3 (2¥C, Ar), 129.0, 127.6, 112.6 (2¥C, Ar), 20.2 (Me); HRMS Calcd for $C_{14}H_{13}N_3O_3Cl_2$: 373.0055, Found: 373.0059.

2-Chloro-5-fluoro-benzoic acid *N*¢**-2-fluorophenyl-hydrazide (1o-Cl).** Yield 78%; mp 145.5 *◦*C; ¹ H NMR (400 MHz, DMSOd6) *d* 10.34 (br s, 1H), 7.89 (br s, 1H), 7.61 (dd, 1H, *J* = 4.9, 8.8 Hz), 7.35–7.46 (m, 2H), 6.96–7.14 (m, 3H), 6.71–6.79 (m,

1H); 13C NMR (136 Hz, DMSO-d6) *d* 165.04 (165.02), 161.6, 159.1, 136.55 (136.47), 136.41 (136.30), 131.84 (131.76), 125.77, (125.74), 124.60 (124.57), 119.05 (118.99), 118.50 (118.29), 116.38 (116.13), 114.97 (114.79), 113.88 (113.85); HRMS Calcd for $C_{13}H_9N_2OClF_2: 282.0371$, Found: 282.0374.

General procedures for the synthesis of indazole-3-ones 2a–o. The mixture of 2-chlorobenzoic acid hydrazide (1.0 mmol), CuI (0.95 mg, 0.005 mmol, 0.5 mol%), L-proline (23 mg, 0.20 mmol, 20 mol%), and K_2CO_3 (0.28 g, 2.0 mmol) in DMSO (10 mL) was stirred at 90 *◦*C for 24 h under nitrogen atmosphere. The mixture was treated with water and the mixture was extracted three times with ethyl acetate. The combined organic layer was washed with water and brine and dried over magnesium sulfate. After filtration, solvent was evaporated *in vacuo* to *ca.* 1 mL and crystallization afforded indazole-3-ones.

1-Phenylindazol-3(2H)-one (2a)^{7,20}. Yield 80%; R_f 0.45 (hexane : AcOEt = 7 : 3); mp 166.4 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) δ 11.29 (br s, 1H, NH), 7.76 (t, 2H, $J = Ar$), 7.68 (d, 2H, *J* = 7.8 Hz, Ar), 7.51 (t, 2H, *J* = 7.8 Hz, Ar), 7.45 (t, 1H, *J* = 7.3 Hz, Ar), 7.25 (t, 1H, *J* = 7.3 Hz, Ar), 7.15 (t, 1H, *J* = 7.3 Hz, Ar); IR (KBr): v 1554 (C=O), 1349 (NO₂), 1301, 1231, 885, 802, $753,695$ cm⁻¹.

A gram scale preparation of 2a. A mixture of 2-chlorobenzoic acid hydrazide **1a** (1.30 g, 5.25 mmol), CuI (5 mg, 0.026 mmol, 0.5 mol%), L-proline (120 mg, 1.05 mmol, 20 mol%), and K_2CO_3 (1.45 g, 10.5 mmol) in DMSO (15 mL) was stirred at 90 *◦*C for 24 h under nitrogen atmosphere. After cooling, the mixture was treated with sat. NaHCO₃ aq. (100 mL) and the mixture was extracted eight times with ethyl acetate (30 mL \times 8). The combined organic layer was washed with water (50 mL \times 3) and brine (50 mL) and dried over magnesium sulfate. After filtration, solvent was evaporated *in vacuo* to afford a crude product which was recrystallized to provide pure **2a** (0.754 g, 68%). The mother liquid was concentrated *in vacuo* and the residue was subjected to column chromatography (chloroform) afforded additional pure **2a** $(0.244 \text{ g}, 22\%)$. HET: "CNMR (139 Hz, DMS0-46) 8 (139 H4 (139 Hz, DMS0-46) 8 February 2012 Published on 17 November 2012 Published on 17 November 2012 Published on 17 November 2012 Published Distributions (149 Bers of 2012 Angers of 2013 P

5-Methyl-1-phenyl-2,3-dihydro-1*H***-indazol-3-one (2b).** Yield 87%; mp 226.5 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 11.16 (br s, 1H), 7.63–7.72 (m, 3H), 7.46–7.56 (m, 3H), 7.29 (d, 1H, *J* = 8.8 Hz), 7.24 (t, 1H, *J* = 7.3 Hz), 2.41 (s, 3H); 13C NMR (136 Hz, DMSOd6) *d* 155.9, 140.4, 137.9, 130.1, 129.4 (2¥C), 129.3, 124.4, 120.1 (2×C), 119.5, 115.1, 110.2, 20.6; HRMS-EI Calcd for $C_{14}H_{12}N_2O$: 224.0949, Found: 224.0938.

6-Nitro-1-phenyl-2,3-dihydro-1*H***-indazol-3-one (2c).** Yield 93%; *R*_f 0.31 (hexane : AcOEt = 7 : 3); mp 286.9 °C; ¹H NMR (400 MHz, DMSO-d6) *d* 11.86 (br s, 1H, NH), 8.46 (s, 1H, Ar), 8.00 (d, 1H, *J* = 8.5 Hz), 7.94 (d, 1H, *J* = 8.5 Hz), 7.74 (d, 2H, *J* = 7.6 Hz, Ar), 7.59 (t, 2H, *J* = 7.6 Hz, Ar), 7.37 (t, 1H, *J* = 7.3 Hz, Ar); ¹³C NMR (136 Hz, DMSO-d6) δ 156.4 (C=O), 147.7, 139.2, 137.8, 130.1 (2¥C, Ar), 126.5, 122.1, 121.8 (2¥C, Ar), 117.8, 114.9,106.8; IR (KBr): v 3093, 1554, 1349, cm⁻¹; HRMS-EI Calcd for $C_{13}H_9N_3O_3$: 255.0644, Found: 255.0605.

6 -Nitro - 1 - (4 -methoxy - phenyl) - 2,3 - dihydro-1*H***-indazol-3-one (2d).** Yield 55%; R_f 0.21 (hexane : AcOEt = 7 : 3); mp 281.4 °C; ¹H NMR (400 MHz, DMSO-d6) *δ* 11.70 (br s, 1H, CONH), 8.30 (s, 1H, Ar), 7.78–8.07 (m, 2H, Ar), 7.61 (d, 2H, *J* = 6.1 Hz, Ar),

7.13 (d, 2H, *J* = 6.3 Hz, Ar), 3.82 (s, 3H, OMe); 13C NMR (136 Hz, DMSO-d6) *δ* 157.8 (C=O), 155.8, 147.3, 137.8, 132.1, 123.7 (2¥C, Ar), 121.8, 116.9, 114.9 (2¥C, Ar), 114.1, 106.4, 55.5 (OMe); HRMS Calcd for C₁₄H₁₁N₃O₄: 285.0749, Found: 285.0748.

6-Chloro-1-phenyl-2,3-dihydro-1*H***-indazol-3-one (2e)¹⁸.** Yield 91%; mp 236.4 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 11.47 (br s, 1H), 7.77 (s, 1H), 7.76 (d, 1H, *J* = 8.5 Hz), 7.67 (d, 2H, *J* = 7.8 Hz), 7.52 (t, 2H, *J* = 7.6 Hz), 7.28 (1H, t, *J* = 7.3 Hz), 7.16 (d, 1H, *J* = 9.3 Hz); 13C NMR (136 Hz, DMSO-d6) *d* 156.2, 139.6, 139.3, 133.6, 129.6 (2¥C), 125.4, 122.1, 121.0 (2¥C), 120.9, 113.6, 109.9.

6-Chloro-1-(4-nitrophenyl)-2,3-dihydro-1*H***-indazol-3-one (2f).** Yield 86%; colorless oil; R_f 0.20 (hexane : AcOEt = 7 : 3); ¹H NMR (400 MHz, DMSO-d6) δ 11.97 (br s, 1H, NH), 8.32 (d, 2H, $J =$ 9.0 Hz, Ar), 8.04 (s, 1H, Ar), 7.95 (d, 2H, *J* = 9.0 Hz, Ar), 7.80 (d, 1H, *J* = 8.5 Hz, Ar), 7.28 (d, 1H, *J* = 8.5 Hz, Ar); 13C NMR (136 Hz, DMSO-d6) δ157.7 (C=O), 144.9, 143.0, 139.4, 134.6, 125.5, 125.4 (2¥C, Ar), 122.4, 119.5 (2¥C, Ar), 115.3, 111.1; HRMS-EI Calcd for $C_{13}H_8N_3O_3Cl$: 289.0254, Found: 289.0275.

1-*tert***-Butyl-6-chloro-2,3-dihydro-1***H***-indazol-3-one (2g).** Yield 83%; mp 179.3 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.80 (br s, 1H, NH), 7.74 (s, 1H), 7.68 (d, 1H, *J* = 8.5 Hz), 7.05 (d, 1H, $J = 8.5$ Hz), 1.67 (s, 9H); ¹³C NMR (136 Hz, DMSO-d6) *d* 152.7, 139.2, 131.6, 121.6, 118.8, 112.6, 111.1, 58.8, 29.3 (3¥C); HRMS-EI Calcd for C₁₁H₁₃N₂OCl: 224.0717, Found: 224.0707.

6-Chloro-1-cyclohexyl-2,3-dihydro-1*H***-indazol-3-one (2h).** Yield 82%; mp 197.0; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.77 (br s, 1H, NH), 7.67 (d, 1H, *J* = 7.6 Hz), 7.59 (d, 1H, *J* = 8.5 Hz), 6.96 (1H, dd, *J* = 1.7, 7.6 Hz), 4.32–4.42 (1H, m, NCH), 1.63–1.88 (m, 7H), 1.38–1.53 (m, 2H), 1.12–1.26 (m, 1H); 13C NMR (136 Hz, DMSO-d6) *d* 153.9, 140.2, 131.9, 121.5, 118.9, 110.8, 108.8, 55.8, 32.0 (2¥C), 25.1, 25.0 (2¥C); HRMS-EI Calcd for C13H15N2OCl: 250.0873, Found: 250.0873.

5-Fluoro-1-(4-methylphenyl)-2,3-dihydro-1*H***-indazol-3-one (2i).** Yield 79%; mp 223.4 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 11.25 (br s, 1H, NH), 7.01 (br s, 1H), 7.43–7.60 (m, 3H), 7.24– 7.28 (m, 3H), 2.34 (s, 3H, Me); 13C NMR (136 Hz, DMSO-d6) *d* 155.8, 137.6, 136.2, 134.4, 129.9 (3¥C), 120.8 (3¥C), 117.3, 111.9, 104.5, 20.5; HRMS-EI Calcd for $C_{14}H_{11}N_2$ OF: 242.0855, Found: 242.0854.

1-*tert***-Butyl-5-fluoro-2,3-dihydro-1***H***-indazol-3-one (2j).** Yield 90%; mp 172.8 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 10.53 (br s, 1H, NH), 7.64 (dd, 1H, *J* = 3.9, 9.3 Hz), 7.31 (d, 1H, *J* = 8.5 Hz), 7.15 (t, 1H, *J* = 9.3 Hz), 1.59 (s, 9H); 13C NMR (136 Hz, DMSO-d6) *d* 156.8, 154.4, 152.6, 136.5, 115.6, 113.4, 104.0, 58.5, 29.3 (3×C); HRMS-EI Calcd for $C_{11}H_{13}N_2$ OF: 208.1012, Found: 208.0984.

6-Chloro-1-phenyl-1*H***,2***H***,3***H***-pyrazolo[3,4-b]pyridin-3-one (2k).** Yield 66%; *R*_f 0.64 (hexane : AcOEt = 1 : 1); mp 236.4 °C; ¹H NMR (400 MHz, DMSO-d6) *d* 11.90 (br s, 1H, CONH), 8.25 (dd, 1H, *J* = 8.4, 1.2 Hz Ar), 8.08 (d, 2H, *J* = 8.8 Hz, Ar), 7.51 (t, 2H, *J* = 7.6 Hz, Ar), 7.29 (dd, 1H, *J* = 8.3, 1.2 Hz, Ar), 7.57 (dt, 1H, *J* = 8.3, 1.3 Hz, Ar);¹³C NMR (136 Hz, DMSO-d6)δ 154.4 (C=O), 150.4, 148.5, 138.9, 133.4, 129.2 (2¥C, Ar), 124.9, 119.4 (2¥C, Ar), 116.8, 106.5; HRMS-EI Calcd for $C_{12}H_8N_3OCl$: 245.0356, Found: 245.0327.

6-Chloro-1-(4-chlorophenyl)-1*H***,2***H***,3***H***-pyrazolo[3,4-b]pyridin-3-one (2l).** Yield 90%; mp 266.6 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 11.99 (br s, 1H, NH), 8.26 (d, 1H, *J* = 8.0 Hz), 8.12 (d, 2H, *J* = 7.6 Hz), 7.57 (d, 2H, *J* = 7.6 Hz), 7.31 (d, 1H, *J* = 8.0 Hz);13C NMR (136 Hz, DMSO-d6) *d* 154.6, 150.5, 148.6, 137.7, 133.5, 129.2 (2¥C), 128.6, 120.6 (2¥C), 117.1, 106.8; HRMS-EI Calcd for $C_{12}H_7N_3OCl_2$: 278.9966, Found: 278.9950.

6 -Chloro - 1 - (4 -methylphenyl) - 1*H***,2***H***,3***H***-pyrazolo[3,4-b]pyridin-3-one (2m).** Yield 77%; mp 222.5 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 11.89 (br s, 1H, NH), 8.26 (d, 1H, *J* = 8.3 Hz), 7.93 (d, 2H, *J* = 8.5 Hz), 7.30 (d, 2H, *J* = 8.5 Hz), 7.25 (d, 1H, *J* = 8.3 Hz), 2.32 (s, 3H, Me); 13C NMR (136 Hz, DMSO-d6) *d* 154.2, 150.3, 148.3, 136.5, 134.1, 133.4, 129.5 (2¥C), 119.5 (2¥C), 116.5, 106.2, 20.5; HRMS-EI Calcd for C₁₃H₁₀N₃OCl: 259.0512, Found: 259.0504.

6-Chloro-5-sulfamoyl-1-(4-methylphenyl)-2,3-dihydro-1*H***-indazol-3-one (2n).** Yield 65%; R_f 0.63 (hexane: AcOEt = 1:3); mp 297.7 *◦*C; ¹ H NMR (400 MHz, DMSO-d6) *d* 11.80 (br s, 1H, CONH), 8.45 (s, 1H, Ar), 7.84 (s, 1H, Ar), 7.58 (d, 2H, *J* = 6.6 Hz, Ar), 7.57 (s, 2H, SO₂NH₂), 7.35 (d, 2H, $J = 8.3$ Hz, Ar), 2.36 (s, 3H, OMe); ¹³C NMR (136 Hz, DMSO-d6) δ: 156.8 (C=O), 139.3, 136.7, 135.6, 133.1, 130.2 (2¥C, Ar), 130.1, 122.5, 121.6 (2¥C, Ar), 112.6, 112.0, 20.6; IR (neat): 3382 (NH), 3277 (NH), 3032 (Ar), 1557 (C=O), 1337 (S=O), 1169, 997, 916, 810 cm⁻¹; HRMS-EI Calcd for $C_{14}H_{12}N_3O_3SC$: 337.0268, Found: 337.0168. Cellston-144-blooploogil-147.847 symmoloid-bloopline. Notes and references

2012 Published on 2012 Published on 17 November 2012 Published Cellston-17 November 2012 Published on the CHI and the CH and the CH angers of the

5-Fluoro-1-(2-fluorophenyl)-2,3-dihydro-1*H***-indazol-3-one (2o)¹³.** Yield 58%; *R*_f 0.35 (hexane : AcOEt = 7 : 3); mp 211.7 [°]C; ¹H NMR $(400 \text{ MHz}, \text{DMSO-46})$ δ 11.39 (br s, 1H, CONH), 7.59 (t, 1H, $J =$ 7.8 Hz Ar), 7.40–7.54 (m, 3H, Ar), 7.36 (t, 1H, *J* = 7.3 Hz, Ar), 7.22–7.34 (m, 2H, Ar).

Pharmacology²¹

Cell Culture. HeLa S3 cells were cultured at 37 *◦*C in a humidified atmosphere with 5% CO₂, in a RPMI 1640 cell culture medium supplemented with 10% fetal bovine serum.

In vitro **cytotoxicity test.** The cells were plated at a density of 1.5×103 cells per 0.1 mL per well in 96-well flat-bottomed microplates. After overnight incubation, 0.1 mL aliquots of medium containing serially diluted test compounds were added to triplicate wells, and incubation was continued for 72 more hours. Cell growth was assessed using the crystal violet staining method.**²²** The IC_{50} value was defined at the drug concentration needed to produce a 50% reduction of growth relative to the untreated control.

Acknowledgements

We gratefully acknowledge Ms. Sachiko Asami, Mrs. Ikuhiro Inada, Atsushi Isayama, and Hiroaki Ohmukai for generous supports in some of the experiments.

Notes and references

- 1 For a review, see: S. Bräse, C. Gil and K. Knepper, *Bioorg. Med. Chem.*, 2002, **10**, 2415.
- 2 S. R. Fletcher, E. Mclver, S. Lewis, F. Burkamp, C. Leech, G. Mason, S. Boyce, D. Morrison, G. Richards, K. Sutton and A. B. Jones, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 2872.
- 3 H. Wang, H. Han and D. D. Von Hoff, *Cancer Res.*, 2006, **66**, 9722.
- 4 N. Kawanishi, T. Sugimoto, J. Shibata, K. Nakamura, K. Masutani, M. Ikuta and H. Hirai, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 5122.
- 5 K. A. M. Abouzid and H. S. El-Abhar, *Arch. Pharmacal Res.*, 2003, **26**, 1.
- 6 (*a*) B. Silvestrini and C. Y. Cheng, *US Pat.* 99-304042, 1999; (*b*) H. Cerecetto, A. Gerpe, M. Gonzalez, V. J. Aran and C. O. de Ocariz, *Mini-Rev. Med. Chem.*, 2005, **5**, 869.
- 7 S. Tanimori, Y. Ozaki, Y. Iesaki and M. Kirihata, *Synlett*, 2008, 1973.
- 8 Recent synthesis of indazolones, see: (*a*) M. B. Donald, W. E. Conrad, S. James, J. S. Oakdale, J. D. Butler, M. J. Haddadin and M. J. Kurth, *Org. Lett.*, 2010, **12**, 2524; (*b*) G. Dou and D. Shi, *J. Comb. Chem.*, 2009, **11**, 1073; (*c*) N. Halland, M. Nazare', O. R'kyek, J. Alonso, M. Urmann and A. Lindenschmidt, *Angew. Chem., Int. Ed.*, 2009, **48**, 6879; (d) D. Viña, E. del Olmo, J. L. López-Pérez and A. San Feliciano, *Org. Lett.*, 2007, **9**, 525; (*e*) W. Stadlbauer, N. Camp, in *Science of Synthesis: Houben-Weyl Methods of Molecular Transformations*, ed. D. Bellus, S. V. Ley, R. Noyori, M. Regitz, E. Schaumann, E. Shinkai, E. J. Thomas, B. M. Trost and P. J. Reider, Thieme, Stuggart, 2002, vol. 12, p 227.
- 9 Recent reviews for copper-catalyzed Ullmann-type coupling reaction, see: (*a*) F. Bellina and R. Rossi, *Chem. Rev.*, 2010, **110**, 1082; (*b*) F. Monnier and M. Taillefer, *Angew. Chem., Int. Ed.*, 2009, **48**, 6954; (*c*) G. Evano, N. Blanchard and M. Toumi, *Chem. Rev.*, 2008, **108**, 3054; (*d*) D. Ma and Q. Cai, *Acc. Chem. Res.*, 2008, **41**, 1450; (*e*) M. Carril, R. SanMartin and E. Dom´ınguez, *Chem. Soc. Rev.*, 2008, **37**, 639; (*f*) M. Kienle, S. R. Dubbaka, K. Brade and P. Knochel, *Eur. J. Org. Chem.*, 2007, 4166 and references cited therein.
- 10 When 10 mol% of L-proline was employed with 10 mol% of CuI, the yield of **2a** was 29% at room temperature for 36 h and 66% at 70 *◦*C for 12 h, respectively.
- 11 The efficiency is even better and comparable to corresponding iodoarene (91% yield; conditions: 10 mol% CuI, 20 mol% L-proline, 2 equiv. K₂CO₃ in DMSO, at 70 [°]C for 3 h) and bromoarene (62^o% yield; conditions: 10 mol% CuI, 20 mol% L-proline, 2 equiv. K_2CO_3 in DMSO, at room temperature for 12 h)**⁷** .
- 12 S. Tanimori, H. Kashiwagi, T. Nishimura and M. Kirihata, *Adv. Synth. Catal.*, 2010, **352**, 2531 See, also: P.-F. Larsson, A. Correa, M. Carril, P.-O. Norrby and C. Bolm, *Angew. Chem., Int. Ed.*, 2009, **48**, 5691.
- 13 J. Magano, M. Waldo, D. Greene and E. Nord, *Org. Process Res. Dev.*, 2008, **12**, 877.
- 14 R. M. Schelkun and P.-W. Yuen, PCT*Int. Appl.*(2006), WO 2006056873 A2 20060601.
- 15 S. Tanimori, T. Nishimura and M. Kirihata, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 4119.
- 16 K. Kitatani, T. Takeda and S. Hoshi, *Jpn. Kokai Tokkyo Koho*, 1994, JP 06027652 A 19940204.
- 17 C.-Y. Zhang, X.-H. Liu, B.-L. Wang, S.-H. Wang and Z.-M. Li, *Chem. Biol. Drug Des.*, 2010, **75**, 489.
- 18 AKos Building Blocks Product List (AKos Consulting and Solutions Deutschland GmbH).
- 19 R. W. Addor, D. G. Kuhn and D. P. Wright Jr., *US* 4814349 A 19890321, 1989.
- 20 D. L. Selwood *et al.*, *J. Med. Chem.*, 2001, **44**, 78.
- 21 Y. Aoyagi, N. Masuko, S. Ohkubo, M. Kitade, K. Nagai, S. Okazaki, K. Wierzba, T. Terada, Y. Sugimoto and Y. Yamada, *Cancer Sci.*, 2005, **96**, 614.
- 22 T. Utsugi, K. Aoyagi, T. Asao, S. Okazaki, Y. Aoyagi, M. Sano, K. Wierzba and Y. Yamada, *Jpn J. Cancer Res.*, 1997, **88**, 992.